



Desmids (Zygnematophyceae) from the littoral zone of an urban artificial lake: taxonomic aspects and geographical distribution

Desmídias (Zygnematophyceae) da zona litorânea de um lago artificial urbano:
aspectos taxonômicos e distribuição geográfica

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Abstract: Aim: The aim of this study was to assess the taxonomic aspects and geographical distribution of the periphytic desmids from two different substrates in the littoral zone of an urban artificial lake. **Methods:** Samples of epiphytic and epilithic desmids were collected by grass and rock scraping, respectively, in two stations in the Cascavel municipal lake, Paraná, during April 2015, and March 2016 and 2017. In the laboratory, the taxa obtained from the samples were photographed and their morphometric characteristics recorded. The species composition of the desmid was characterized, and the frequency of occurrence as well as the geographic distribution of each taxon in the Paraná State were recorded. **Results:** We identified 35 taxa, distributed in 10 genera. The most representative were *Closterium* Nitzsch *ex* Ralfs and *Cosmarium* Corda *ex* Ralfs. A high occurrence of sporadic taxa (60%) was observed and among the recorded taxa, the most frequent were *Cosmarium pseudoconnatum* Nordstedt var. *pseudoconnatum* and *Desmidiium grevillei* (Kützing *ex* Ralfs) De Bary. Nine taxa were exclusive to the epilithic substrate and only *Netrium digitus* (Ralfs) Itzigsohn & Rothe var. *digitus* was exclusive to the epiphytic substrate, while the remaining 25 taxa occurred in both substrates. Seven taxa are new records for the Paraná State. **Conclusion:** Although our results are of qualitative nature, they provide baseline data on the biogeographic distribution of taxa, which are crucial for future taxonomic and ecological studies that might contribute to the preservation of the biodiversity.

Keywords: aquatic biodiversity; Desmidiales; geographic distribution; lentic environment; periphytic algae.

Resumo: Objetivo: O objetivo deste estudo foi avaliar os aspectos taxonômicos e a distribuição geográfica das desmídias perifíticas de dois substratos diferentes na zona litorânea de um lago artificial urbano. **Métodos:** Amostras de desmídias epífitas e epilíticas foram coletadas por raspagem de



gramíneas e rochas, respectivamente, em duas estações no lago municipal de Cascavel, Paraná, durante abril de 2015 e março de 2016 e 2017. No laboratório, os táxons obtidos a partir das amostras foram fotografados e suas características morfométricas foram registradas. A composição das espécies de desmídias foi caracterizada e a frequência de ocorrência, bem como a distribuição geográfica de cada táxon no Estado do Paraná foram registradas. **Resultados:** Nós identificamos 35 táxons, distribuídos em 10 gêneros. Os gêneros mais representativos foram *Closterium* Nitzsch ex Ralfs e *Cosmarium* Corda ex Ralfs. Uma alta ocorrência de táxons esporádicos (60%) foi observada, e entre os táxons registrados, os mais frequentes foram *Cosmarium pseudoconnatum* Nordstedt var. *pseudoconnatum* e *Desmidiium grevillei* (Kützinger ex Ralfs) De Bary. Nove táxons foram exclusivos para o substrato epilítico e somente *Netrium digitus* (Ralfs) Itzigsohn & Rothe var. *digitus* foi exclusivo para o substrato epifítico, enquanto os demais 25 táxons ocorreram em ambos os substratos. Sete táxons são novas ocorrências para o estado do Paraná. **Conclusão:** Embora nossos resultados sejam de natureza qualitativa, eles fornecem uma base de dados sobre a distribuição biogeográfica dos táxons, os quais são cruciais para futuros estudos taxonômicos e ecológicos que podem contribuir para a preservação da biodiversidade.

Palavras-chave: algas perifíticas; ambiente lêntico; biodiversidade aquática; Desmídias; distribuição geográfica.

1. Introduction

Landscapes are composed of different aquatic ecosystems. In these ecosystems, conditions of light and nutrients are appropriate for the development and establishment of microorganisms that are key elements for their functioning. Artificial lakes are remarkable components of the local and regional landscapes, in where a wide, but a still little-known diversity of microalgae can be found (Menezes et al., 2011a).

The periphyton is a complex assemblage of different microorganisms, including microalgae, cyanobacteria, microinvertebrates, and detritus attached to organic or inorganic substrates (Wetzel, 1983). Depending on the type of substrate to which they are attached, they are known as epilithon, if attached to rocky substrates, and epiphyton, if attached to aquatic plants (Wetzel, 1981; Franceschini et al., 2010). Periphytic microalgae play key roles in the energy and matter cycles in the littoral zones (Vercellino & Bicudo, 2006), being their temporal and spatial dynamics influenced by environmental conditions (Moschini-Carlos, 1999; Goldsborough & Robinson, 1996).

The desmids belong to the class Zygnematophyceae (Phylum Charophyta) and are considered the greatest and most diverse group of the Infrakingdom Streptophyta (Gontcharov & Melkonian, 2005; Ruggiero et al., 2015). They are important components of the structure of the periphyton community, due to the high diversity of taxa and of functional strategies (Felisberto et al., 2014).

Desmids are cosmopolitan and *k* strategists that exhibit a wide variety of forms, being either unicellular or pseudofilamentous (Coesel, 1996). They are considered the geographically most

studied group of algae due to their often appealing appearance (Coesel & Krienitz, 2008). Desmids have a few truly planktonic representatives, which are mostly associated with periphyton, in mesotrophic to oligotrophic environments of slightly acid pH, high temperatures and low electrical conductivity (Coesel, 1982, 1996; Černá & Neustupa, 2010; Stamenković & Hanelt, 2017). In addition, desmids are ecologically sensitive organisms to environmental changes and thus can be useful tools as bioindicators for the monitoring and management of aquatic environments (Coesel, 2001). For these reasons, taxonomic and floristic inventories, often ignored, become relevant for understanding the complex association of these organisms with the environment, and can contribute to the better knowledge of the biodiversity and the consequences of its changes (Meester & Declerck, 2005).

The aim of this study was to assess the epilithic and epiphytic composition of desmids in the littoral zone of an urban artificial lake, as well as the geographic distribution of the taxa recorded, contributing to the knowledge on the desmid diversity of the Paraná State.

2. Material and Methods

2.1. Study area

The Cascavel River belongs to the Cascavel River basin and is the main water source of the municipality of Cascavel, Paraná. Its principal springs are located in the municipal lake region (24°32' and 25°17' S and 53°05' and 53°50' W), which is placed in the Paulo Gorski Ecological Park (Casagrande, 1996). The Cascavel municipal lake (Figure 1) is supplied by several streams and has a drainage area of 117.5 km². This lake is an

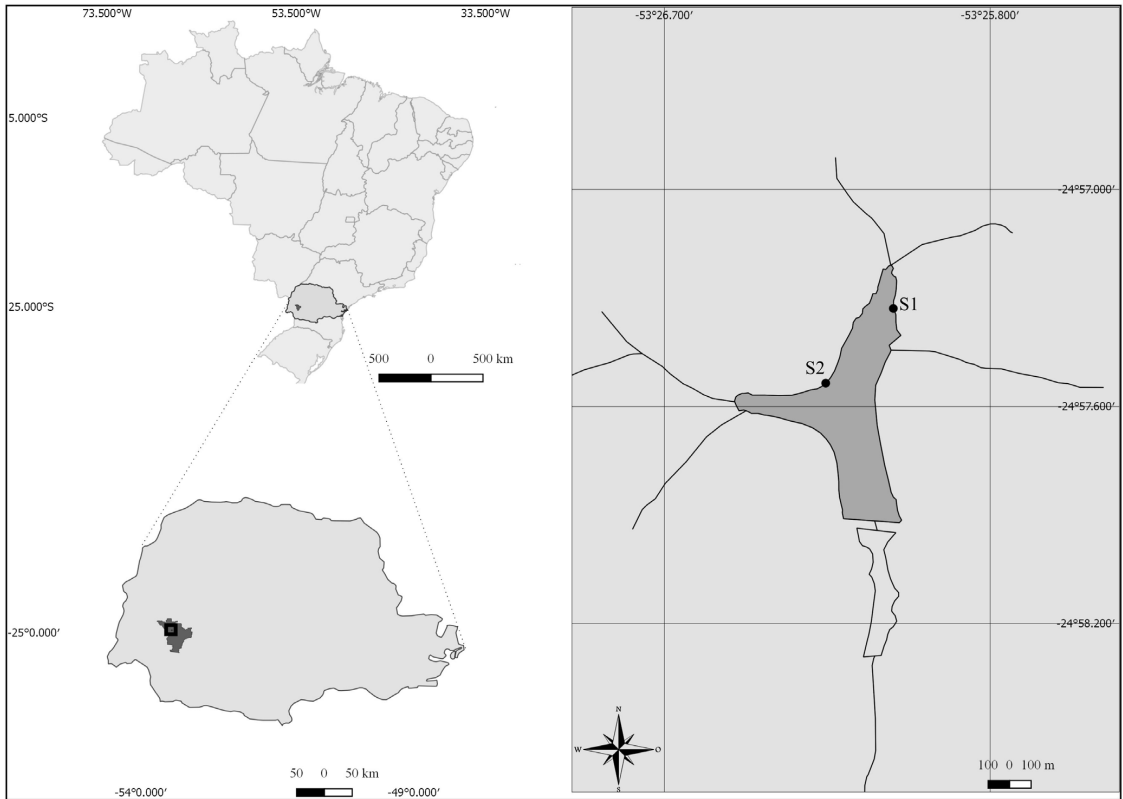


Figure 1. Location of the sampling stations in the Cascavel municipal lake, Paraná, Brazil (S1 – Station 1; S2 – Station 2).

artificial reservoir intended to the public supply and constitutes an important area of touristic visitation. During the last 20 years, the lake has been suffering from sedimentation of its margins, besides sewage discharges in its surroundings. This region has a humid-mesothermal subtropical climate, with a well-defined summer and winter periods and well-distributed rainfalls through the year (Alvares et al., 2014). Two sampling stations (S1: 24°57'19.73" S, 53°26'3.97" W and S2: 24°57'32.11" S, 52°26'15.22" W) were established in the lake. At each station, some environmental variables were measured with a Horiba U-50 multiparameter probe (Figure 1; Table 1).

2.2. Sampling and laboratory analysis

Collection of samples and measurement of environmental variables at each sampling station were performed during April 2015, and March 2016 and 2017 (n = 12). Biological samples were collected from epiphytic and epilithic substrates in the littoral zone of the lake, by grass and rock scraping, respectively. Samples were properly stored and preserved in 1:1 *Transeau* solution (Bicudo &

Table 1. Characterization of the sampling stations in the Cascavel municipal lake, Paraná, Brazil. Values of the abiotic variables are expressed as means and coefficients of variation (%).

	WT	pH	DO	EC	TDS	TUR
S1	19.3 (17%)	6.6 (4%)	9.4 (5%)	0.05 (10%)	0.03 (12%)	0.93 (62%)
S2	21.3 (22%)	6.4 (5%)	10.1 (16%)	0.05 (49%)	0.03 (48%)	28.40 (12%)

References: WT – water temperature, °C; pH; DO – dissolved oxygen, mg L⁻¹; EC – electrical conductivity, μS cm⁻¹; TDS – total dissolved solids, g L⁻¹; TUR – turbidity, NTU; S1 – station 1; S2 – station 2.

Menezes, 2006). For qualitative analyses, an average of 10 temporary slides per sample was prepared, and the microalgae photographed and characterized morphometrically under 400 or 1000x magnification with an Olympus trinocular microscope model CX31 with camera attached.

The systematic framework of the genera followed Bicudo & Menezes (2006). Identification of *Cosmarium* and *Actinotaenium* was according to Prescott et al. (1981), and specialized scientific papers

were used for other genera. Taxa were characterized morphometrically and their geographic distribution in the Paraná State recorded. Measurements were expressed in µm and represented by the symbols L: (length), W: (width), and I: (isthmus). The samples were deposited in the Herbarium of the State University of West Paraná (UNOP-Algae), Cascavel campus (Table 2).

The frequency of occurrence of the taxa in the samples (constancy = C) was calculated according to Dajoz (2005). Taxa were classified as constant (C ≥ 70%), common (30% ≤ C ≤ 70%), sporadic (10% ≥ C ≤ 30%) or rare (C ≤ 10%). Artificial keys to identify the taxa in each genus were constructed for those genera more than two taxa.

3. Results and Discussion

We recorded 35 taxa of desmids, distributed in 10 genera: *Actinotaenium* (Nägeli) Teiling (1), *Closterium* Nitzsch ex Ralfs (10), *Cosmarium* Corda ex Ralfs (12), *Desmidium* C. Agardh ex Ralfs (1), *Euastrum* Ehrenberg ex Ralfs (2), *Micrasterias* C. Agardh ex Ralfs (3), *Netrium* (Nägeli) Itzigsohn & Rothe (2), *Penium* Brébisson ex Ralfs (2), *Pleurotaenium* Nägeli (1) and *Staurastrum* Meyen ex Ralfs (1). All recorded taxa are described below.

Actinotaenium cucurbitinum var. ***cucurbitinum*** (Bisset) Teiling f. ***minus*** (West & G. S. West) Teiling, Bot. Not., 4: 376-426. 1954.

Basionym: *Penium cucurbitinum* f. *minus* West & G. S. West, 1894.

Cells 2.2-2.8 times longer than wide. Semicells cylindrical, slightly attenuated toward the apex. Apex rounded. Median constriction with shallow sinus. Cell wall finely punctuated. Chloroplast with lamellae. One pyrenoid per

semicell. L: 40.4-63.1 µm; W: 14.4-27.6 µm; I: 13.7-26.7 µm. Figure 2

Geographic distribution in the Paraná State: First record of the taxon.

Frequency of occurrence: Common

Identification key for the taxa recorded in the genus *Closterium*

1. Cells slightly curved, almost straight or straight2
2. Cell wall with fine longitudinal striae.....3
3. Apex rounded or truncated-rounded.....4
4. Pyrenoids 12-14
.....*Closterium acerosum* var. *acerosum*
4. Pyrenoids 4-6.....
.....*Closterium kuetzingii* var. *kuetzingii*
3. Apex truncated.....5
5. Cell length 216-233 µm
.....*Closterium bailyanum* var. *bailyanum*
5. Cell length 867.6 µm.....
.....*Closterium braunii*
2. Cell wall smooth6
6. Cell length 344.9-388 µm
.....*Closterium pseudolunula* var. *pseudolunula*
6. Cell length 33.6-213.4 µm.....7
7. Chloroplast axial with 4-5 longitudinal lamellae and pyrenoids 1-3
.....*Closterium navicula* var. *navicula*
7. Chloroplast axial divided transversally and pyrenoids 3-5
.....*Closterium closterioides* var. *closterioides*
1. Cells lunate, curved or strongly curved8
8. Apex truncated
.....*Closterium tumidum* var. *tumidum*
8. Apex rounded or rounded-conical9

Table 2. Reference information of the examined samples collected in the Cascavel municipal lake and deposited in the Herbarium of the State University of West Paraná (UNOP-Algae), Cascavel, Paraná, Brazil.

UNOP-Algae	Collection date	Station	Geographical Coordinates	Collector
4490	01/03/2017	S1	24°57'19.73" S 53° 26'3.97" W	N.C. Bueno
4491	01/03/2017	S1	24°57'19.73" S 53° 26'3.97" W	N.C. Bueno
4492	01/03/2017	S2	24°57'32.11" S 52°26'15.22" W	N.C. Bueno
4493	01/03/2017	S2	24°57'32.11" S 52°26'15.22" W	N.C. Bueno
2617	05/03/2016	S1	24°57'19.73" S 53° 26'3.97" W	N.C. Bueno
2618	05/03/2016	S1	24°57'19.73" S 53° 26'3.97" W	N.C. Bueno
2615	05/03/2016	S2	24°57'32.11" S 52°26'15.22" W	N.C. Bueno
2616	05/03/2016	S2	24°57'32.11" S 52°26'15.22" W	N.C. Bueno
4187	04/04/2015	S1	24°57'19.73" S 53° 26'3.97" W	N.C. Bueno
4188	04/04/2015	S1	24°57'19.73" S 53° 26'3.97" W	N.C. Bueno
4185	04/04/2015	S2	24°57'32.11" S 52°26'15.22" W	N.C. Bueno
4186	04/04/2015	S2	24°57'32.11" S 52°26'15.22" W	N.C. Bueno

9. Cell wall with striae.....
*Closterium malmei* var. *malmei*
 9. Cell wall smooth
*Closterium incurvum* var. *incurvum*.

Closterium acerosum Ehrenberg ex Ralfs var. ***acerosum***, British Desmidiaceae 164, pl. 27, figs a,b,d-e. 1848.

Cells 11-17 times longer than wide. Cells slightly lunate, with ventral margin slightly concave and dorsal margins slightly convex, tapering towards both ends. Apex truncated-rounded, often angular. Cell wall colorless or brownish striated. Chloroplast axial. Pyrenoids 12-14. L: 493.8-598.4 µm; W: 29-50.3 µm. Figures 3-5

Geographic distribution in the Paraná State: First record of the taxon.

Frequency of occurrence: Sporadic

Closterium baillyanum (Brébisson ex Ralfs) Brébisson var. ***baillyanum***, Mém. Soc. Imp. Sc. Nat. Cherbourg, 4: 151, pl. 1-2. 1856.

Basionym: *Closterium didymotocum* Ralfs var. *baillyanum* Brébisson in Ralfs Brit. Desm., 169. 1848.

Cells 6-10 times longer than wide. Cells slightly curved with dorsal margin slightly convex; ventral margin a bit concave, sometimes more straight in the median portion. Apex truncated. Cell wall with fine striae and dense punctuations, mainly in the apical region. Chloroplast axial. Pyrenoids 5-8, arranged in a median series. L: 216-233.6 µm; W: 23.8-35.9 µm. Figure 6

Geographic distribution in the Paraná State: Bortolini et al. (2009); Menezes et al. (2011a); Aquino et al. (2014).

Frequency of occurrence: Sporadic

Closterium braunii Reinsch, Abh. Senck. Naturf. Ges., 6 (2): 138, pl. 20C, fig. 1: 1-5. 1867.

Cells 12 times longer than wide. Cells slightly curved with dorsal margin convex and ventral margin a bit concave. Apex truncated. Cell wall finely striated. Chloroplast axial. Pyrenoids 12-14. L: 867.6 µm; W: 69.3 µm. Figure 7-9.

Geographic distribution in the Paraná State: First record of the taxon.

Frequency of occurrence: Rare

Closterium closterioides (Ralfs) A. Louis & Peeters var. ***closterioides***, Bull. Jard. Bot. Natl. Belg., 37: 410, pl. 13, fig. 119. 1967.

Basionym: *Penium closterioides* Ralfs 152, pl. 34: fig. 4. 1848.

Cells 3-4 times longer than wide. Cells straight, elliptic to fusiform, with both dorsal and ventral margins equally convex. Apex truncated-rounded.

Cell wall smooth, colorless. Chloroplast axial divided incompletely and transversally. Pyrenoids 3-5, arranged in a median series. L: 91.9-142.6 µm; W: 23.8-57.5 µm. Figure 10.

Geographic distribution in the Paraná State: Algarte et al. (2006); Felisberto & Rodrigues (2007, 2012); Bortolini et al. (2009, 2010a); Aquino et al. (2014); Burack et al. (2016).

Frequency of occurrence: Sporadic

Closterium incurvum Brébisson var. ***incurvum***, Mém. Soc. Imp. Sc. Nat. Cherbourg, 4: 150, pl. 2, fig. 47. 1856.

Cells 2-4 times longer than wide. Cells lunate, strongly curved, dorsal margin convex and ventral margin concave, with pointed-rounded poles. Cell wall smooth or hyaline to brownish. Chloroplast axial. Pyrenoids 4-7, arranged in a median series. L: 33.6-72.5 µm; W: 15-18.3 µm. Figure 11.

Geographic distribution in the Paraná State: Picelli-Vicentim (1984); Bittencourt-Oliveira & Castro (1993); Felisberto & Rodrigues (2007); Biolo et al. (2008); Bortolini et al. (2009).

Frequency of occurrence: Common

Closterium kuetzingii Brébisson var. ***kuetzingii***, Mém. Soc. Imp. Sc. Nat. Cherbourg, 4: 156, pl. 2, fig. 40. 1856.

Cells 15-22 times longer than wide. Cells almost straight; median portion fusiform with dorsal and ventral margins equally convex. Apex rounded and slightly curved. Cell wall finely striated. Chloroplast axial. Pyrenoids 4-6, arranged in a median series. L: 224.5-391.8 µm; W: 14.2-17.2 µm. Figure 12.

Geographic distribution in the Paraná State: Andrade & Rachou (1954); Bittencourt-Oliveira & Castro (1993); Cecy et al. (1997); Algarte et al. (2006); Menezes et al. (2011a); Aquino et al. (2014).

Frequency of occurrence: Common

Closterium malmei Borge var. ***malmei***, Ark. Bot., 1: 79, pl. 1, fig. 21. 1903.

Cells 2-4 times longer than wide. Cells strongly curved, not inflated in the median region. Apex rounded conical with a thickening in the interior wall. Cell wall colorless to brownish with longitudinal striae. Chloroplast axial. Pyrenoids 9, arranged in a median series. L: 91.3-213.4 µm; W: 32.9-47.9 µm. Figure 13-15.

Geographic distribution in the Paraná State: Algarte et al. (2006); Aquino et al. (2014).

Frequency of occurrence: Sporadic

Closterium navicula (Brébisson) J. Lütkenmüller var. ***navicula***, Beitr. Biol. Pfl. Breslau, 8(3): 395, 405, 408. 1902.

Basionym: *Penium navicula* Brébisson 146, pl 2, fig. 37. 1856.

Cells 3-4 times longer than wide. Cells straight, elliptic to fusiform, with both dorsal and ventral margins equally convex and truncated-rounded apex. Cell wall smooth, colorless. Chloroplast axial with 4-5 longitudinal lamellae. Pyrenoids 1-3, arranged in a median series. L: 34.4-52.6 µm; W: 11.6-15.3 µm. Figure 16.

Geographic distribution in the Paraná State: Algarte et al. (2006); Felisberto & Rodrigues (2007); Bortolini et al. (2009); Menezes et al. (2011a); Aquino et al. (2014); Felisberto et al. (2014); Burack et al. (2016).

Frequency of occurrence: Common

Closterium pseudolunula Borge var. ***pseudolunula***, Ark. Bot., 8 (13): 3, pl. 1, fig. 2. 1909.

Cells 6-8 times longer than wide. Cells straight, elliptic to fusiform with dorsal margin slightly convex and ventral margin straight. Apex truncated-rounded. Cell wall smooth. Chloroplast axial. Pyrenoids not observed. L: 344.9-388 µm; W: 40-57.4 µm. Figure 17-19.

Geographic distribution in the Paraná State: Bittencourt-Oliveira (1993).

Frequency of occurrence: Sporadic

Closterium tumidum L. N. Johnson var. ***tumidum***, Bull. Torrey Bot. Club, 22: 291, pl. 232, fig. 4, 31. 1895.

Basionym: *Closterium cornu* Ehrenberg ex Ralfs 176, pl. 30: figs. 6f- g. 1848.

Cells 5 times longer than wide. Cells curved with dorsal margins convex and ventral margin slightly straight; truncated apex. Cell wall smooth, yellowish or brownish. Chloroplast axial. Pyrenoids 3-5, arranged in a median series. L: 72.6-80.3 µm; W: 13.5-14.8 µm. Figure 20.

Geographic distribution in the Paraná State: Ferreira et al. (2011); Aquino et al. (2014).

Frequency of occurrence: Sporadic

Identification key for the taxa recorded in the genus *Cosmarium*

1. Cell wall smooth or finely punctuated2
2. Cell wall smooth
.....*Cosmarium obsoletum* var. *obsoletum*
2. Cell wall finely punctuated3
3. Apex rounded or rounded-truncated4
4. Shallow median constriction
.....*Cosmarium pseudoconnatum* var. *pseudoconnatum*
4. Deep median constriction5

5. Pyramidal semicells
.....*Cosmarium pseudopyramidatum* var. *excavatum*
5. Subtrapeziform semicells
Cosmarium candianum var. *candianum*
3. Apex truncated6
6. Circular semicell
.....*Cosmarium dispersum* var. *dispersum*
6. Trilobular semicell
Cosmarium trilobulatum var. *trilobulatum*
1. Cell wall with granules, spines or warts7
7. Apex retuse to straight
.....*Cosmarium quadrum* var. *minus*
7. Apex rounded and truncated-rounded8
8. Cell wall with spines
.....*Cosmarium denticulatum* var. *ovale*
8. Cell wall with granules9
9. Cell length 43.8-49 µm
.....*Cosmarium subspeciosum* var. *subspeciosum*
9. Cell length 73.7-92 µm10
10. Cell face ornamented with triangular granules
.....*Cosmarium decoratum*
10. Cell face ornamented without granules11
11. Cells oblong-elliptic
Cosmarium margaritatum var. *margaritatum* f. *margaritatum*
11. Cells pyramidal-truncated
.....*Cosmarium subspeciosum* var. *validius*

Cosmarium candianum Delponte var. ***candianum***, Mém. R. Accad. Sci. Torino, 28: 113, pl. 8, fig. 1-6. 1877.

Cells once longer than wide. Semicells semicircular with rounded lateral and apical margins. Apex rounded. Cell wall finely and closely punctate. Median constriction deep. Linear median sinus closed. Chloroplast axial. Pyrenoids 2 per semicell. L: 45-50 µm; W: 43.3-48 µm; I: 14.5-17 µm. Figure 21.

Geographic distribution in the Paraná State: Picelli-Vicentim (1984); Bortolini et al. (2010b); Menezes et al. (2013); Aquino et al. (2014, 2016).

Frequency of occurrence: Sporadic

Cosmarium decoratum West & G.S. West, Trans. Linn. Soc. London, Bot., II, 5(2): 61, pl. 7, fig. 21. 1895.

Cells 1.3 times longer than wide. Semicells semicircular to pyramidal truncated. Apex rounded-truncated. Cell wall granular, with hexagonal series arranged in dense granules, with



Figures 2-23. Desmids of the Cascavel municipal lake, Paraná, Brazil. **2.** *Actinotaenium cucurbitinum* var. *cucurbitinum* f. *minus* **3-5.** *Closterium acerosum* var. *acerosum* **4-5.** Detail of the cell wall **6.** *C. baillyanum* var. *baillyanum* **7-9.** *C. braunii* **8-9.** Detail of the apex **10.** *C. closterioides* var. *closterioides* **11.** *C. incurvum* var. *incurvum* **12.** *C. kuetszingii* var. *kuetszingii* **13-15.** *C. malmei* var. *malmei* **14.** Detail of the Apex **15.** Detail of the cell wall **16.** *C. navicula* var. *navicula* **17-19.** *C. pseudolunula* var. *pseudolunula* **18.** Detail of the Apex **19.** Detail of the cell wall **20.** *C. tumidum* var. *tumidum* **21.** *Cosmarium candianum* var. *candianum* **22.** *C. decoratum* **23.** *C. denticulatum* var. *ovale*. Scale 10 µm.

triangular pores in the central region. Median constriction deep. Sinus slightly open in the proximal region. Chloroplast axial. Pyrenoids 2 per semicell. L: 73.7-85.5 µm; W: 55-64.6 µm; I: 24.4-28.4 µm. Figure 22.

Geographic distribution in the Paraná State: Felisberto & Rodrigues (2010a); Biolo et al. (2013); Aquino et al. (2014, 2016).

Frequency of occurrence: Sporadic

Cosmarium denticulatum Borge var. *ovale* Grönblad, Acta Soc. Sci. fenn.: sér. B, 2(6): 17, pl. 5, fig. 103. 1945.

Cells 1.6 times longer than wide. Semicells pyramidal with lateral and apical margins denticulate; lateral margins convex, sometimes slightly straight, with rounded-truncated apex. Cell wall with spines. Median constriction deep. Linear median sinus closed. Chloroplast axial. Pyrenoids not observed. L: 139 µm; W: 83.7 µm; I: 31 µm. Figure 23.

Geographic distribution in the Paraná State: Felisberto & Rodrigues (2010a); Bortolini et al. (2010b); Aquino et al. (2014).

Frequency of occurrence: Rare

Cosmarium dispersum L. N. Johnson f. *dispersum* L. N. Johnson, Bull. Torrey Bot. Club, 22: 297, pl. 233, fig. 19. 1895.

Cells about as long as wide. Semicells approximately circular, with truncated apex. Cell wall finely punctuated, showing approximately 18 undulations in the semicell margins. Median constriction deep. Linear sinus closed. Chloroplast axial. Pyrenoids 2 per semicell. L: 45.2-47.5 µm; W: 39.5-42.2 µm; I: 14.6-14.8 µm. Figure 24.

Geographic distribution in the Paraná State: Aquino et al. (2014, 2016).

Frequency of occurrence: Sporadic

Cosmarium margaritatum (P. Lundell) Roy & Bisset var. *margaritatum* f. *margaritatum*, Jd. Bot., 27(1): 194. 1886.

Basionym: *Cosmarium latum* Brébisson var. *margaritatum* P. Lundell, Nova Acta R. Soc. Scient. Upsal.: sér. 3, 8(2): 26. 1871.

Cells 1.2 times longer than wide. Semicells subretangular, sometimes ellipsoidal-subretangular, with rounded angles. Apex straight or very slightly convex. Cell wall uniformly granulate, with 28 to 32 solid granules distributed on the margins of the semicell. Median constriction deep, linear median sinus. Chloroplast axial. Pyrenoids 2 per semicell. L: 78.6 µm; W: 64 µm; I: 21.3 µm. Figure 25-26.

Geographic distribution in the Paraná State: Lozovei & Luz (1976); Lozovei & Hohmann (1977); Cecy et al. (1997); Cetto et al. (2004); Silva & Cecy (2004); Bortolini et al. (2010b); Menezes et al. (2011b); Aquino et al. (2014, 2016).

Frequency of occurrence: Rare

Cosmarium obsoletum (Hantzsch) Reinsch var. *obsoletum*, Acta Soc. Senckenberg, 6: 142, pl. 22D1, fig. 1-4. 1867.

Basionym: *Arthrodesmus obsoletus* Hantzsch in Rabenhorst, Algen Europa's. n° 1407. 1862

Cells 1.1 times wider than long. Semicells semicircular with rounded margins and slightly truncated apex. Cell wall smooth. Median constriction deep with closed linear sinus, slightly open at the extremities. Chloroplast axial. Pyrenoids 2 per semicell. L: 43.1 µm; W: 48.5 µm; I: 17 µm. Figure 27.

Geographic distribution in the Paraná State: Aquino et al. (2016).

Frequency of occurrence: Rare

Cosmarium pseudoconnatum Nordstedt var. *pseudoconnatum*, Vidensk. Medd. Naturh. For. Kjöbenhavn, 21: 214, pl. 3, fig. 17. 1870.

Cells 1.2-1.3 times longer than wide. Semicells semicircular with rounded-oval outline and smooth margins. Apex rounded. Cell wall finely punctuated. Median constriction shallow. Median sinus open. Chloroplast axial, radiate. Pyrenoids 2 per semicell. L: 41.9-60 µm; W: 31-49.4 µm; I: 25-55 µm. Figure 28-29.

Geographic distribution in the Paraná State: Bittencourt-Oliveira (1993); Cecy et al. (1997); Picelli-Vicentim et al. (2001); Silva & Cecy (2004); Felisberto & Rodrigues (2005a, 2008); Bortolini et al. (2010b); Menezes et al. (2011b); Aquino et al. (2014).

Frequency of occurrence: Common

Cosmarium pseudopyramidatum P. Lundell var. *excavatum* (Nordstedt) Krieger & Gerloff, Die Gattung *Cosmarium*, Lief 2: 127, taf 26, fig. 7. 1965.

Basionym: *Cosmarium pseudopyramidatum* P. Lundell f. *excavata* Nordstedt., Acta Univ. Lund., 19: 13. 1873.

Cells 1.6 times longer than wide. Semicells pyramidal-truncated, basal angles obliquely rounded; lateral margins widely concave, convergent towards the apex. Apex truncated or rounded. Cell wall punctate. Median constriction deep. Linear sinus closed. Chloroplast axial. Pyrenoids not observed. L: 62.5-75.3 µm; W: 37.5-45.7 µm; I: 16.5-19.9 µm. Figure 30.

Geographic distribution in the Paraná State: First record of the taxon.

Frequency of occurrence: Sporadic

Cosmarium quadrum P. Lundell var. *minus* Nordstedt, Lund. Univ. Arsskr., 9 (10): 11. 1873.

Cells slightly wider than long. Semicells subrectangular with lateral and apical margins undulate, slightly convex and straight. Apex retuse to straight. Cell wall ornamented with warts arranged in vertical rows, following the cell's outline. Median constriction deep. Linear median sinus closed, with dilated at the extremities. Chloroplast axial. Pyrenoids 2 per semicell. L: 37.3-39.2 µm; W: 39.6-40.7 µm; I: 12.4-12.5 µm. Figure 31.

Geographic distribution in the Paraná State: Felisberto & Rodrigues (2005a, b, 2012); Bortolini et al. (2010b); Menezes et al. (2011b); Aquino et al. (2014, 2016).

Frequency of occurrence: Sporadic

Cosmarium subspeciosum Nordstedt var. *subspeciosum*, Öfv., Kongl. Vet.-Akad. Förhandl., 32(6): 22, pl. 6, fig. 13. 1875.

Cells 1.3-1.4 times longer than wide. Semicells semicircular to pyramidal, with crenulate margins. Apex truncated-rounded, granulate. Cell wall densely granulate; granules in radial series toward the cellular center, median region with some angles. Median constriction deep. Linear median sinus closed. Chloroplast axial. Pyrenoids 2 per semicell. L: 43.8-49 µm; W: 30-35.8 µm; I: 10.1-13.3 µm. Figure 32.

Geographic distribution in the Paraná State: Felisberto & Rodrigues (2005a, 2008); Bortolini et al. (2010a); Menezes et al. (2011b); Aquino et al. (2014, 2016).

Frequency of occurrence: Sporadic

Cosmarium subspeciosum Nordstedt var. *validius* Nordstedt, K. Svenska Vet.-Akad. Handl., 22 (8): 49, pl. 5, fig. 10. 1888.

Cell 1.4 times longer than wide. Semicells pyramidal-truncated with lateral and apical margins with prominent crenulations, and truncate-rounded apex. Cell wall granulate, granules in radial series forming lines toward the region center, central protuberance with vertical lines of granules. Median constriction deep. Linear median sinus closed. Basal angles abruptly rounded. Chloroplast axial. Pyrenoids 2. L: 77.5-92 µm; W: 52.5-62.5 µm; I: 17-19.4 µm. Figure 33.

Geographic distribution in the Paraná State: Silva & Cecy (2004); Felisberto & Rodrigues (2005a, 2008); Bortolini et al. (2010b); Aquino et al. (2014).

Frequency of occurrence: Sporadic

Cosmarium trilobulatum Reinsch var. *trilobulatum*, Acta Soc. Senckenberg, 6: 118(10), pl. 22(3) A II, fig. 1-6. 1867.

Cell 1.1-1.2 times longer than wide. Semicell subtrapeziform, 3-lobed with slightly convex lateral margins. Apex wide, truncated. Cell wall smooth or finely punctate. Median constriction deep, with linear sinus closed. Chloroplast axial. Pyrenoids 1 per semicell. L: 25.8-27.6 µm; W: 20.5-28 µm; I: 7.8-10 µm. Figure 34.

Geographic distribution in the Paraná State: Felisberto & Rodrigues (2005a, b, 2010a, 2012); Algarte et al. (2006); Biolo & Rodrigues (2011); Aquino et al. (2014).

Frequency of occurrence: Sporadic

Desmidium grevillei (Kützing ex Ralfs) De Bary, Ein Beit. phys. besch. Botanik., 76, pl. 4, fig. 30-31. 1858.

Basionym: *Didymoprium grevillea* Kützing ex Ralfs 57, pl. 2. 1848.

Cell 2.1-2.3 times wider than long. Semicells oblong with rounded lateral margins. Filament not twisted. Cell wall smooth. Median constriction shallow. Sinus open. Chloroplast axial. Pyrenoids not observed. L: 17.5-22.9 µm; W: 41.2-48.2 µm; I: 40-47 µm. Figure 35.

Geographic distribution in the Paraná State: Bittencourt-Oliveira (1993); Bortolini et al. (2008); Felisberto & Rodrigues (2010b); Menezes et al. (2011a, 2013).

Frequency of occurrence: Common

Identification key for the taxa recorded in the genus *Euastrum*

1. Face of semicell ornamented with 3 mucilage pores.....*Euastrum* sp.

1. Face of semicell without mucilage pores.....*Euastrum bidentatum* var. *bidentatum*

Euastrum bidentatum Nägeli var. *bidentatum*, Gatt. Einz. Algen, 122, pl. 7D, fig. 1a-f. 1849.

Cell 1.3-1.4 times longer than wide. Semicells semi-elliptical. Apical margin truncate-rounded, angular and with a wide V-shaped incision, ornamented with a spine-like process. Median constriction deep, with closed linear sinus. Face of semicell ornamented with several granules. Chloroplast axial. Pyrenoids 1 per semicell. L: 47.1-47.2 µm; W: 33.2-34.9 µm; I: 7.5-7.8 µm. Figure 36-37.

Geographic distribution in the Paraná State: First record of the taxon.

Frequency of occurrence: Sporadic

Euastrum sp.

Cell 2 times longer than wide. Semicells semi-elliptical. Apical margin truncated with a wide incision. Median constriction deep, linear sinus closed.

Face of semicell ornamented with 3 mucilage pores. Chloroplast axial. Pyrenoids 1 per semicell. L: 61.4-90 µm; W: 30.5-44.3 µm; I: 10.1-15µm. Figure 38-39.

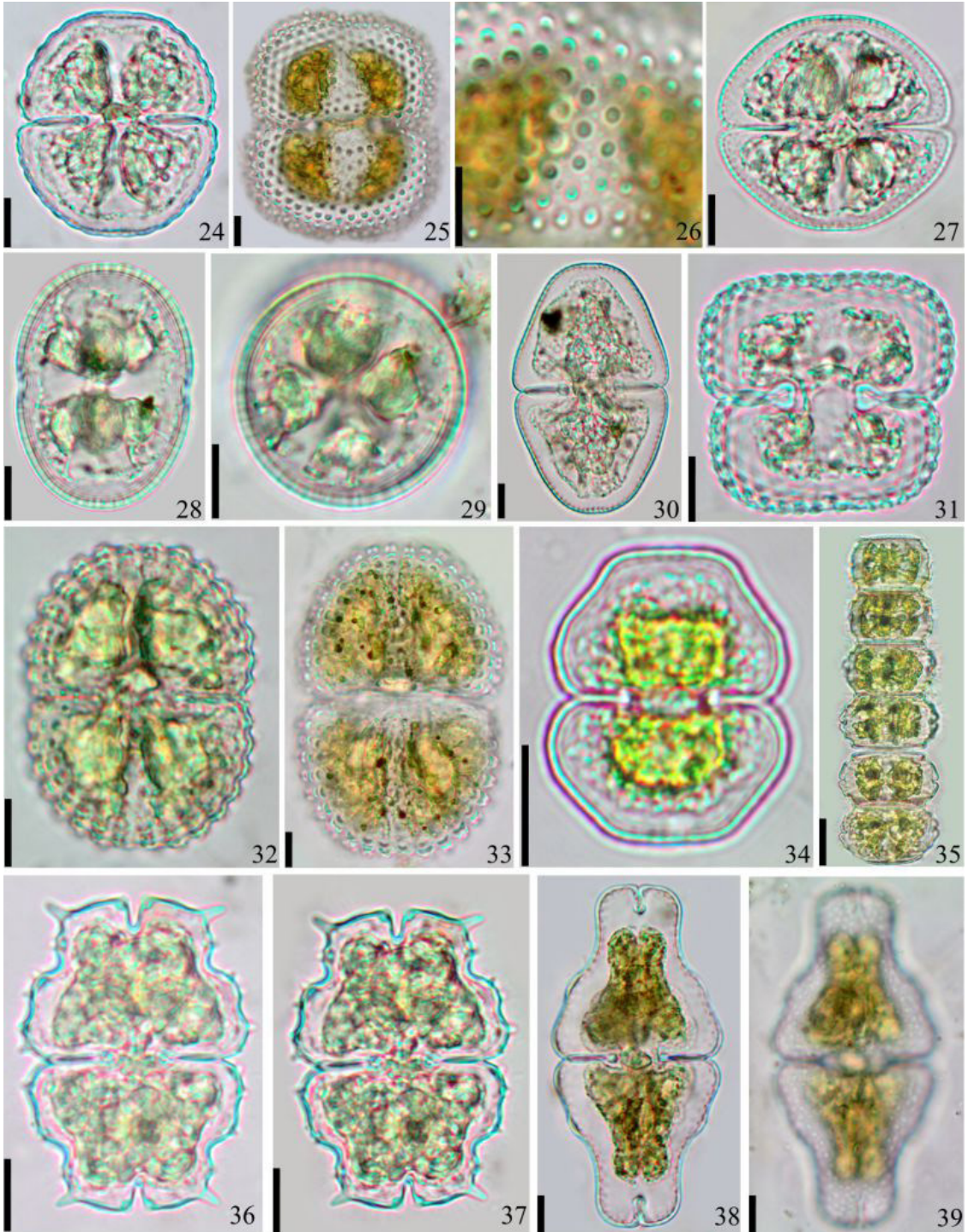


Figure 24-39. Desmids of the Cascavel municipal lake, Paraná, Brazil. 24. *Cosmarium dispersum* f. *dispersum* 25-26. *C. margaritatum* var. *margaritatum* f. *margaritatum* 26. Detail of the cell wall 27. *C. obsoletum* var. *obsoletum* 28-29. *C. pseudoconnatum* var. *pseudoconnatum* 29. Apical view 30. *C. pseudopyramidatum* var. *excavatum* 31. *C. quadrum* var. *minus*. 32. *C. subspeciosum* var. *subspeciosum* 33. *C. subspeciosum* var. *validius* 34. *C. trilobulatum* var. *trilobulatum* 35. *Desmidium grevillei* 36-37. *Euastrum bidentatum* var. *bidentatum* 37. Detail of the cell wall 38-39. *Euastrum* sp. 39. Detail of the cell wall. Scale 10 µm.

Geographic distribution in the Paraná State:
First record of the taxon.

Frequency of occurrence: Sporadic

Identification key for the taxa recorded in the genus *Micrasterias*

1. Semicells 3-lobed.....*Micrasterias laticeps* var. *laticeps*

1. Semicells 5-lobed.....2

2. Cell wall punctuated

.....*Micrasterias quadridentata*

2. Cell wall ornamented with short spines

.....*Micrasterias borgei* var. *borgei*

Micrasterias borgei Krieger var. ***borgei*** in Rabenhorst Kryptogamen-Fl. Deutschl., 13 (2): 86, pl.128, fig. 1-4. 1939.

Cells 1.0-1.1 times longer than wide. Semicells elliptic to elliptical-oblong, 5-lobed, with acutangular interlobular incisions. Apical lobe long, narrow, extending slightly besides the lateral lobes. Cell wall ornamented with short spines. Median constriction deep. Linear sinus closed, slightly open at the extremities. Lateral and basal lobes separated by a linear incision, acutangular at the distal portion. Basal lobes subdivided in 2-4 lobules, acutangular, 2-4 denticulate. Lateral lobes subdivided in 4-8 lobules, acutangular. Chloroplast axial. Numerous pyrenoids. L: 235-265.7 µm; W: 214.8-230 µm; I: 29.6-47.5 µm. Figure 40-42.

Geographic distribution in the Paraná State: Bittencourt-Oliveira & Mecnas (1994); Algarte et al. (2006); Felisberto & Rodrigues (2005a, 2011); Moresco et al. (2009); Menezes et al. (2011a).

Frequency of occurrence: Common

Micrasterias laticeps Nordstedt var. ***laticeps***, Vidensk. Meddr Naturh. Foren., (1869), 14-15: 220. 1870.

Cells 1.2-1.3 times wider than long or 1.0 times longer than wide. Semicells elliptic, 3-lobed with superior margin convex to straight in the median portion. Apical lobe fusiform to subcuneiform. Interlobular incisions acutangular. Cell wall finely punctuated. Median constriction deep. Basal lobe semifusiform, with 2-denticulated extremities. Chloroplast axial. Multiple pyrenoids. L: 110-176.8 µm; W: 127.6-227.6 µm; I: 20.8-40.3 µm. Figure 43-44.

Geographic distribution in the Paraná State: Andrade & Rachou (1954); Lozovei & Hohmann (1977); Picelli-Vicentim (1984); Bittencourt-Oliveira & Mecnas (1994); Bittencourt-Oliveira (2002); Felisberto & Rodrigues (2005a, 2008, 2010b, 2011); Biolo et al. (2008).

Frequency of occurrence: Sporadic

Micrasterias quadridentata (Nordstedt) Grönblad, Acta Soc. Fauna Flora Fenn., 47(4): 36. 1920.

Cell 1.1-1.2 times longer than wide. Semicell circular, 5-lobed. Interlobular incisions linear, narrow. Apical lobe long, narrow, not extending besides the lateral lobes, margins straight or slightly concave, opening to the beveled apex. Angles acuminate or 2-denticulated. Cell wall punctuated. Median constriction deep. Sinus linear. Basal and lateral lobes subdivided in 4 lobules by 3 incisions. Apex of lobules truncated, sometimes beveled in the median portion, angles with 1 denticle. Chloroplast axial. Multiple pyrenoids. L: 271.3-465 µm; W: 229-374.7 µm; I: 33.1-60 µm. Figure 45.

Geographic distribution in the Paraná State: Moresco et al. (2009); Menezes et al. (2011a).

Frequency of occurrence: Common

Identification key for the taxa recorded for the genus *Netrium*

1. Chloroplast with lamellae and lateral projections.....*Netrium digitus* var. *lamellosum*

1. Chloroplast without lamellae or projections.....*Netrium digitus* var. *digitus*

Netrium digitus (Ralfs) Itzigsohn & Rothe var. ***digitus*** in Rabenhorst, Alg. Sachsen, 508. 1856.

Basionym: *Penium digitus* Ralfs, Brit. Desmidiaceae. 150, pl. 25, fig. 3. 1848

Cells 3-4 times longer than wide. Cells oblong-elliptical, with lateral margins slightly convex. Apex truncated-rounded. Cell wall smooth. Chloroplast axial with denticulate projections. Pyrenoids 2-3, longitudinally distributed. L: 121.9-266.4 µm; W: 39.1-67.5 µm. Figure 46.

Geographic distribution in the Paraná State: Felisberto & Rodrigues (2008); Bortolini et al. (2010a).

Frequency of occurrence: Sporadic

Netrium digitus (Ehrenberg ex Ralfs) Itzigsohn & Rothe var. ***lamellosum*** (Brébisson) Grönblad, Acta Soc. Fauna Flora Fenn., 47(4): 13. 1920.

Basionym: *Penium lamellosum* Bréb., Mém. Soc. Sci. Nat. Cherbourg 4: 146, pl. 2, fig. 34. 1856.

Cells 5-6 times longer than wide. Cells narrow, elliptical-elongate, with lateral subparallel margins. Apex rounded. Cell wall smooth. Median constriction shallow. Chloroplast axial, with prominent lateral projections. Pyrenoids 2-3, arranged in the median region of the chloroplast. L: 270.5-342.7 µm; W: 49-51.8 µm. Figure 47.

Geographic distribution in the Paraná State: Felisberto & Rodrigues (2008).

Frequency of occurrence: Sporadic

Identification key for the taxa recorded in the genus *Penium*

1. Cell wall with longitudinal spiral striae of uneven spaced granules*Penium spirostriolatum*

1. Cell wall with longitudinal irregular lines of small granules.....*Penium margaritaceum* var. *margaritaceum*

Penium margaritaceum (Ehrenberg) Brébisson ex Ralfs var. *margaritaceum*, Brit. Desm., 149, pl. 25, fig. 3; pl. 33, fig. 1. 1848.

Basionym: *Closterium margaritaceum* Ehrenberg, 95, pl. 6, fig. 13. 1838.

Cell 4-5 times longer than wide. Semicells elongate-cylindrical, with parallel margins. Apex subtruncated-rounded. Cell wall red-brownish, ornamented with longitudinal irregular lines of small granules; presence of elongation zones. Pyrenoids 1, spherical and central in each branch. L: 95.5-132 µm; W: 20.2-22.1 µm. Figure 48-49.

Geographic distribution in the Paraná State: Felisberto & Rodrigues (2005a, 2010b); Biolo et al. (2008); Aquino et al. (2014).

Frequency of occurrence: Sporadic

Penium spirostriolatum J. Barker, Quart. J. Microscop. Sci., series 2, 9: 194. 1869.

Cell 6 times longer than wide. Semicells cylindrical and elongate. Apex truncated-rounded. Cell wall brownish, ornamented with longitudinal spiral striae of uneven spaced granules; presence of several constrictions along the cell. Median constriction shallow or little apparent. Chloroplast 2-4 per cell, each one with 5-7 longitudinal rings. Pyrenoids central, spherical to slightly elongate in each branch. L: 125.7 µm, W: 18.9 µm. Figure 50.

Geographic distribution in the Paraná State: First record of the taxon.

Frequency of occurrence: Rare

Pleurotaenium trabecula (Ehrenberg) Nägeli var. *trabecula* f. *trabecula*, Gatt. Einzell. Alg., 104, pl.6, fig. A. 1849.

Basionym: *Closterium trabecula* Ehrenberg, Infusoria, 62: 70. 1830.

Cell 9-15 times longer than wide. Semicells cylindrical with lateral margins parallel; 1 basal intumescence, 1-2 light undulations above the intumescence; rounded angles. Apical margin retuse, smooth and with subquadratic angles. Cell wall hyaline, punctuated. Median constriction shallow. Median sinus open. Chloroplast parietal, in the form of a longitudinal band. Several pyrenoids per chloroplast. L: 533-561 µm, W: 34.6-60 µm; I: 28.6-40 µm. Figure 51.

Geographic distribution in the Paraná State: Cetto et al. (2004); Bortolini et al. (2008); Aquino et al. (2014).

Frequency of occurrence: Sporadic

Staurastrum alternans Brébisson ex Ralfs var. *alternans*, Brit. Desm., 132, pl. 21, fig. 7. 1848.

Cell 1.2 times longer than wide. Semicell triangular, oblong with rounded angles. Short process. Apex flattened, truncated. Cell wall granulated. Median constriction deep. Sinus open. Chloroplast axial. Pyrenoids 1 per semicell. L: 23.2-26.7 µm; W: 18-21.8 µm; I: 8.4-8.9 µm. Figure 52-53.

Geographic distribution in the Paraná State: Bittencourt-Oliveira & Mecnas (1994); Felisberto & Rodrigues (2008, 2013).

Frequency of occurrence: Common

In this study, even though we did not attempt to make any quantitative estimation, a decrease in the number of taxa was recognized throughout the analyzed period. The greatest number of taxa was recorded in 2015, when 33 taxa occurred in the epilithic substrate and 18 taxa in the epiphytic substrate, while in 2016 and 2017 the number of taxa recorded in both substrates was smaller (17 and 5, respectively). Among the taxa recorded, nine were exclusive to the epilithic substrate and only *Netrium digitus* (Ralfs) Itzigsohn & Rothe var. *digitus* was recovered from the epiphytic substrate, whereas the other 25 taxa occurred on both substrates (Table 3). The presence of natural substrates potentially influences the desmids, whose development depends on the type of substrates available (Coesel, 1982; Barbosa et al., 2013). Among the taxa recorded, seven are new records for the Paraná State: *Actinotaenium cucurbitinum* var. *cucurbitinum* (Bisset) Teiling f. *minus*, *Closterium acerosum* (Schrank) Ehrenberg var. *acerosum*, *Closterium braunii* Reinsch, *Cosmarium pseudopyramidatum* P. Lundel var. *excavatum* Börgesen, *Euastrum bidentatum* Nägeli var. *bidentatum*, *Euastrum* sp. and *Penium spirostriolatum* J. Barke.

The genera *Cosmarium* (12 taxa) and *Closterium* (10 taxa) presented the greatest number of taxa. These genera are cosmopolitan and exhibit high diversity in different environmental scenarios (Coesel, 1996), being common in oligotrophic and mesotrophic aquatic systems (Graham & Wilcox, 2000). Members of *Cosmarium* are ubiquitous and have developed numerous protective strategies (e.g. low number of thylakoids per granum, long and exposed stroma lamellae,

numerous and moderately large plastoglobules and large starch grains) to survive higher temperature conditions at various latitudes

and altitudes (Stamenković & Hanelt, 2017). Regarding the frequency of occurrence, most taxa were sporadic, followed by common and

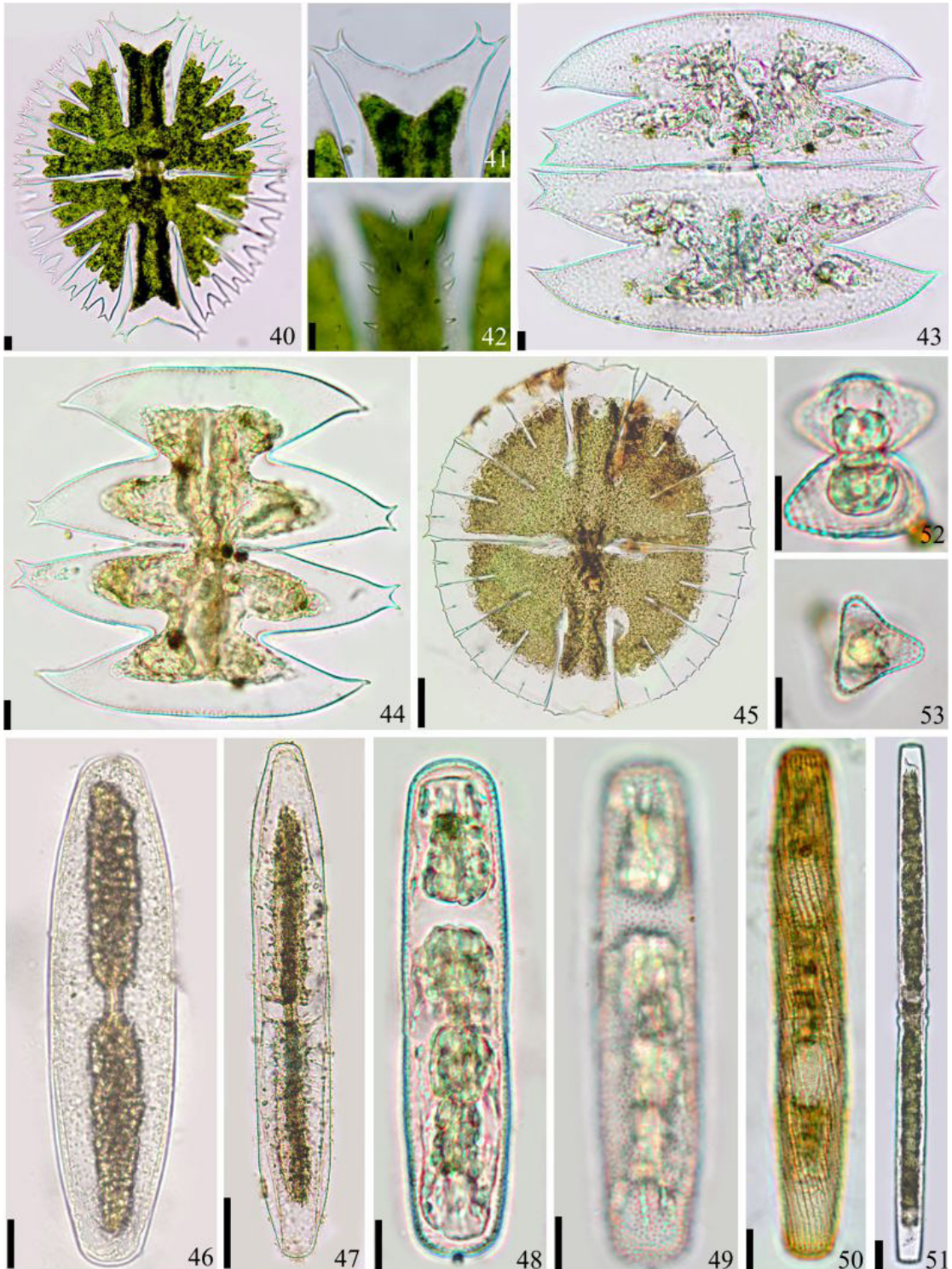


Figure 40-53. Desmids of the Cascavel municipal lake, Paraná, Brazil. **40-42.** *Micrasterias borgei* var. *borgei* **41.** Detail of the Apex **42.** Detail of the cell wall **43-44.** *M. laticeps* var. *laticeps* **45.** *M. quadridentata* **46.** *Netrium digitus* var. *digitus* **47.** *N. digitus* var. *lamellosum* **48-49.** *Penium margaritaceum* var. *margaritaceum* **49.** Detail of the cell wall **50.** *P. spirostriolatum* **51.** *Pleurotaenium trabecula* var. *trabecula* f. *trabecula* **52-53.** *Staurastrum alternans* var. *alternans* **53.** Apical view. Scale 10 μ m.

rare taxa, while no constant taxa were recorded (Table 3). *Cosmarium pseudoconnatum* Nordstedt var. *pseudoconnatum* and *Desmidiium grevillei* (Kützing ex Ralfs) De Bary were the taxa with the greatest occurrence (58% in both cases).

Although the results obtained in the present study are of qualitative nature, they provide baseline data on the biogeographic distribution of taxa in continental aquatic environments. Desmids are considered as excellent indicators of water quality

Table 3. List of occurrence of the periphytic desmids taxa in the Cascavel municipal lake in different substrates (EP – epilithon; EF – epiphyton), in the sampling performed in 2015, 2016 and 2017 (FO - frequency of occurrence, C – Common, S – Sporadic, R – Rare). Emphasizing the occurrence (●) or absence (-) of taxa.

Taxa	2015		2016		2017		FO	Voucher UNOP-Algae
	EP	EF	EP	EF	EP	EF		
<i>Actinotaenium cucurbitinum</i> var. <i>cucurbitinum</i> (Bisset) Teiling f. <i>minus</i> (West & G.S. West) Teiling	●	●	-	●	-	-	C	2615, 2616, 4186, 4187, 4188
<i>Closterium acerosum</i> Ehrenberg ex Ralfs var. <i>acerosum</i>	●	●	-	-	-	-	S	4185, 4187, 4188
<i>C. baillyanum</i> (Brébisson ex Ralfs) Brébisson var. <i>baillyanum</i>	●	●	-	-	-	-	S	4185, 4186, 4188
<i>C. braunii</i> Reinsch	●	-	-	-	-	-	R	4187
<i>C. closterioides</i> (Ralfs) A. Louis & Peeters var. <i>closterioides</i>	●	●	-	-	-	-	S	4186, 4188
<i>C. incurvum</i> Brébisson var. <i>incurvum</i>	●	●	-	●	-	-	C	4185, 4186, 4188, 2615, 2616
<i>C. kuetzingii</i> Brébisson var. <i>kuetzingii</i>	●	-	-	●	-	-	C	2616, 4186, 4187, 4188
<i>C. malmei</i> Borge var. <i>malmei</i>	●	●	-	-	-	-	S	4185, 4188
<i>C. navicula</i> (Brébisson) J. Lütkenmüller var. <i>navicula</i>	●	●	●	●	-	-	C	2615, 2616, 2618, 4185, 4188
<i>C. pseudolunula</i> Borge var. <i>pseudolunula</i>	●	-	-	-	-	-	S	4187, 4188
<i>C. tumidum</i> L.N. Johnson var. <i>tumidum</i>	-	-	●	-	-	-	S	2617, 2618
<i>Cosmarium candianum</i> Delponte var. <i>candianum</i>	●	●	-	-	-	-	S	4186, 4187, 4188
<i>C. decoratum</i> West & G.S. West	●	●	-	-	-	-	S	4185, 4186, 4188
<i>C. denticulatum</i> Borge var. <i>ovale</i> Grönblad	●	-	-	-	-	-	R	4188
<i>C. dispersum</i> L.N. Johnson f. <i>dispersum</i> L.N. Johnson	●	-	-	●	-	-	S	2615, 4188
<i>C. margaritatum</i> (P. Lundell) Roy & Bisset var. <i>margaritatum</i> f. <i>margaritatum</i>	●	-	-	-	-	-	R	4188
<i>C. obsoletum</i> (Hantzsch) Reinsch var. <i>obsoletum</i>	●	-	-	-	-	-	R	4188
<i>C. pseudoconnatum</i> Nordstedt var. <i>pseudoconnatum</i>	●	●	●	●	-	-	C	2615, 2616, 2618, 4185, 4186, 4187, 4188
<i>C. pseudopyramidatum</i> P. Lundell var. <i>excavatum</i> (Nordstedt) Krieger & Gerloff	●	●	-	-	-	-	S	4185, 4186, 4188
<i>C. quadrum</i> Lundell var. <i>minus</i> Nordstedt	●	-	-	-	-	●	S	4187, 4188, 4493
<i>C. subspicosum</i> Nordstedt. var. <i>subspicosum</i>	●	-	-	●	-	-	S	2615, 2616, 4188
<i>C. subspicosum</i> Nordstedt. var. <i>validius</i> Nordstedt	●	●	-	-	-	-	S	4186, 4188
<i>C. trilobulatum</i> Reinsch var. <i>trilobulatum</i>	●	-	-	●	-	-	S	2615, 2616, 4188
<i>Desmidiium grevillei</i> (Kützing ex Ralfs) De Bary	●	-	●	●	●	●	C	2617, 4185, 4186, 4187, 4188, 4491, 4492
<i>Euastrum bidentatum</i> Nägeli var. <i>bidentatum</i>	●	-	-	-	-	-	S	4187, 4188
<i>Euastrum</i> sp.	●	●	-	-	-	-	S	4186, 4187, 4188
<i>Micrasterias borgei</i> Krieger var. <i>borgei</i>	●	●	-	-	●	-	C	4185, 4186, 4188, 4491
<i>M. laticeps</i> Nordstedt var. <i>laticeps</i>	●	-	-	-	-	-	S	4187, 4188
<i>M. quadridentata</i> (Nordstedt) Grönblad	●	●	-	-	●	-	C	4185, 4187, 4188, 4491
<i>Netrium digitus</i> (Ralfs) Itzigsohn & Rothe var. <i>digitus</i>	-	●	-	●	-	-	S	2615, 4185, 4186
<i>N. digitus</i> (Ehrenberg ex Ralfs) Itzigsohn & Rothe var. <i>lamellosum</i> (Brébisson) Grönblad	●	●	-	-	-	-	S	4185, 4186, 4188
<i>Penium margaritaceum</i> (Ehrenberg) Brébisson ex Ralfs var. <i>margaritaceum</i>	●	-	-	●	-	-	S	2615, 2616, 4188
<i>P. spirostriolatum</i> J. Barker	●	-	-	-	-	-	R	4188
<i>Pleurotaenium trabecula</i> (Ehrenberg) Nägeli var. <i>trabecula</i> f. <i>trabecula</i>	●	●	-	-	-	-	S	4186, 4187, 4188
<i>Staurastrum alternans</i> Brébisson ex Ralfs var. <i>alternans</i>	●	-	●	●	-	-	C	2615, 2616, 2618, 4187

because they are highly sensitive to changes in the environmental conditions. Therefore, studies of this nature are of great importance for the monitoring of aquatic ecosystems, once the local and regional biodiversity knowledge provides the base to propose measures of preservation and conservation of the biodiversity.

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